

## BRIEF COMMUNICATIONS

### COMPOSITIONAL HETEROGENEITY OF SULFATED POLYSACCHARIDES SYNTHESIZED BY THE BROWN ALGA *Costaria costata*

T. I. Imbs,<sup>1\*</sup> N. M. Shevchenko,<sup>1</sup> T. L. Semenova,<sup>2</sup>  
S. V. Sukhoverkhov,<sup>2</sup> and T. N. Zvyagintseva<sup>1</sup>

UDC 547.458; 582.272

Brown algae contain sulfated polysaccharides, fucoidans, that exhibit various biological activities [1]. The variety of activities exhibited by fucoidans is related to their structural variations. It was shown that species of a single family (genus) of brown algae can contain sulfated polysaccharides that differ in structure and biological activity [2]. Correspondingly, several different fractions of fucoidans can be isolated from a single species [3, 4]. Our goal was to determine the composition of sulfated polysaccharides synthesized by the brown alga *Costaria costata* [Turn.] Saund (Laminariaceae), which is broadly distributed in seas of the Russian Far East.

Polysaccharides were isolated by acid extraction at room temperature [5] from specimens of *C. costata* collected in July in Troits Bay (Sea of Japan). The polysaccharide fraction was separated into fucoidan (F) and laminaran by chromatography over the hydrophobic sorbent Polikhrom-1 [5]. The monosaccharide composition of the acid-hydrolysis products of the polysaccharides was determined by HPLC in an IC-5000 Biotronik carbohydrate analyzer (Shim-pack ISA-07/S2504 column, 0.4 × 25 cm, potassium borate buffer, flow rate 0.6 mL/min). Monosaccharides were detected using the bicinchoninate method. Monosaccharides (Rha, Man, Fuc, Gal, Xyl, Glc) were used as standards. The molecular weights (MWs) of the polysaccharides were determined by HPLC in a Shimadzu LC-20A instrument with an RID-10A refractometric detector. Fucoidan samples were dissolved in doubly distilled water and filtered through a membrane filter (0.45 µm pore size). Fucoidans were separated over successively connected columns of Shodex Asahipak GS-520 HQ and GS-620 HQ (7.5 × 300 mm) at 50°C with elution by H<sub>2</sub>O (0.8 mL/min). Columns were calibrated using standard pullulans of MWs from 180 to 667,000 Da (Polymer Laboratories, USA) and blue dextran (Amersham, Sweden).

Analysis of the monosaccharide composition of fraction F showed that fucose and galactose were the major monomers. Mannose, rhamnose, xylose, and glucose were detected in smaller quantities. The MW distribution of fraction F was heterogeneous. Three maxima with the strongest at 300 kDa and weaker ones at 80 and 560 were observed in the MW distribution curve.

Fucoidan from *C. costata* collected in July had the following characteristics: yield 2.5% (% of dry alga); uronic acids, 2.2% (of fraction weight); protein, 3.1% (of fraction weight); SO<sub>3</sub>Na<sup>-</sup>, 17.3% (of fraction weight); monosaccharides (mol%): Fuc, 55.1; Gal, 18.1; Man, 9.2; Rha, 11.5; Xyl, 4.3; Glc, 1.8. Protein was determined by the Lowry method [6]; sulfates, by turbidimetry [7]; uronic acids, spectrophotometrically [8]. Fucoidan (F) was also fractionated by ion-exchange chromatography over DEAE-cellulose (3.5 × 14 cm). Retained polysaccharides were eluted by a linear gradient of H<sub>2</sub>O–NaCl (2 M) to afford fractions F-0.5 and F-1.5 (Table 1).

Fraction F-0.5 contained a low-sulfated fucoidan with a heterogeneous monosaccharide composition and a high content of mannose and uronic acids (37 and 15.5%, respectively, of total monosaccharides) (Table 1). Uronic acid in F-0.5 and F-1.5 was identified after total hydrolysis as glucuronic acid using GC of the polyol acetates [9]. This fraction was characterized as uronofucomannans according to the monosaccharide composition determined using GC of the polyol acetates [9].

1) Pacific Institute of Bioorganic Chemistry, Far-East Branch, Russian Academy of Sciences, 690022, Vladivostok, fax: (4232) 31 40 50; e-mail: technolog@piboc.dvo.ru; 2) Institute of Chemistry, Far-East Branch, Russian Academy of Sciences, 690022, Vladivostok, pr. 100-Letiya Vladivostoka, 159. Translated from *Khimiya Prirodnnykh Soedinenii*, No. 1, pp. 86–87, January–February, 2011. Original article submitted June 1, 2010.

Table 1. Characteristics of Fractions F-0.5 and F-1.5 Obtained by DEAE-Cellulose Chromatography of Fucoidan from *C. costata*

Fraction	Yield, %**	MW <sub>av</sub> , kDa	SO <sub>3</sub> Na <sup>-</sup> , %*	Monosaccharides, mol%						
				Fuc	Gal	Man	Rha	Xyl	Glc	GlcA
F-0.5	29	80, 620	6.7	22.6	6.4	37.0	4.2	4.6	9.6	15.5
F-1.5	69	300, 80	23.8	70.2	19.8	7.0	0.0	0.0	0.0	3.0

\*% of fucoidan fraction weight; \*\*% of initial fraction.

The MW distribution of F-0.5 was heterogeneous. It contained in approximately equal amounts fucoidan of average MW (MW<sub>av</sub>) 80 kDa and 620 kDa.

Fraction F-1.5 consisted of about 70% of the initial F and had a simpler monosaccharide composition than F-0.5. Fucose and galactose (1.0:0.28 ratio) accounted for about 90% of the total monosaccharides. The fraction had a high content of sulfates (Table 1). It was identified as a galactofucan according to the monosaccharide composition. The MW distribution of F-1.5 was also heterogeneous. The principal part consisted of fucoidans with MW<sub>av</sub> 300 kDa with a smaller fraction with MW<sub>av</sub> 80 kDa.

Thus, F isolated from *C. costata* collected in July contained as a minimum two groups of fucose-containing polysaccharides with different structures in a 1:2 ratio. These were low-sulfated uronofucomannan with maxima on the MW distribution at about 80 and 600 kDa and high-sulfated galactofucan with MW<sub>av</sub> 300 kDa. The study of the structural characteristics of F fractions isolated from *C. costata* collected in July is continuing.

## ACKNOWLEDGMENT

The work was supported by the RAS Presidium Basic Research Program “Molecular and Cellular Biology”.

## REFERENCES

1. A. I. Usov and M. I. Bilan, *Usp. Khim.*, **78**, No. 8, 846 (2009).
2. A. Cumashi, N. A. Ushakova, M. E. Preobrazhenskaya, A. D'Incecco, A. Piccoli, L. Totani, N. Tinari, G. E. Morozovich, A. E. Berman, M. I. Bilan, A. I. Usov, N. E. Ustyuzhanina, A. A. Grachev, C. J. Sanderson, M. Kelly, G. A. Rabinovich, S. Iacobelli, and N. E. Nifantiev, *Glycobiology*, **17**, 541 (2007).
3. T. Nishino, C. Nishioka, H. Ura, and T. Nagumo, *Carbohydr. Res.*, **255**, 213 (1994).
4. N. M. A. Ponce, C. A. Pujol, E. B. Damonte, M. L. Flores, and C. A. Stortz, *Carbohydr. Res.*, **338**, 153 (2003).
5. T. I. Imbs, N. M. Shevchenko, S. V. Sukhoverkhov, T. L. Semenova, A. V. Skriptsova, and T. N. Zvyagintseva, *Khim. Prir. Soedin.*, 661 (2009).
6. O. H. Lowry, N. J. Rosebrough, A. L. Farr, and R. I. Randall, *J. Biol. Chem.*, **193**, 265 (1951).
7. K. S. Dodgson, *Biochem. J.*, **78**, 312 (1961).
8. N. Blumenkrantz and G. Asboe-Hansen, *Anal. Biochem.*, **54**, 484 (1973).
9. M. Thomas, J. Albersheim, and P. Albersheim, *Plant Physiol.*, **49**, 926 (1972).